A THIOCHOLINE SURFACTANT: PREPARATION AND KINETIC PROPERTIES \*
Robert A. Moss, George O. Bizzigotti, Thomas J. Lukas, Winfred J. Sanders Wright and Rieman Chemistry Laboratories, Department of Chemistry Rutgers, The State University of New Jersey New Brunswick, New Jersey 08903

The great nucleophilicity of thiolate anions and the substantial acidity of thiols have made thiol-functionalized surfactant micelles objectives of intense interest,<sup>1-3</sup> both as esterolysis reagents and as analogs of the cysteine proteinases papain and ficin.<sup>4</sup> Initially, "comicellar" reagents were investigated: <u>N</u>-dodecanoyl-<u>D</u>,<u>L</u>-cysteine,<sup>1</sup> alkane thiols,<sup>2</sup> coenzyme A,<sup>3</sup> and glutathione,<sup>3</sup> each solubilized in micellar alkyltrimethylammonium bromides, accelerated the basic cleavage of <u>P</u>-nitrophenyl acetate (PNPA). More recently, we examined the first selfcontained, thiol-functionalized surfactant, AS-Cys (I).<sup>5</sup> Although micellar AS-Cys is an excellent reagent for the cleavage of PNPA, previous experience with the imidazole-functionalized surfactants, AS-His-Boc (II)<sup>6</sup> and 16-Im (III),<sup>7</sup> suggested that the unknown thiocholine surfactant IV (16-SH) would be even more reactive than AS-Cys. We now report the preparation of 16-SH and its kinetic behavior with PNPA, where 16-SH proves to be the most reactive self-contained functional micellar reagent yet reported.<sup>8</sup>

Choline surfactant V (16-OH) was converted to its triflate, VI (2 equiv.  $Tf_20$ , 1 equiv.  $C_5H_5N$ ,  $CH_2Cl_2$  solution,  $25^\circ$ , 30 min),<sup>9</sup> and the  $CH_2Cl_2$  solution of VI was stirred with excess 1.35 <u>N</u> aqueous sodium thioacetate ( $25^\circ$ , 1 hr), dried and stripped, affording VII (16-SAc, X=OTf, mp 60-64°, 81% yield after 3 recryst. from  $CH_2Cl_2/ether$ ).<sup>10,11</sup> Ion exchange with Dowex 1-X8 (Cl<sup>-</sup> form, 25-50 mesh, H<sub>2</sub>O, 85-90°, 10 min), followed by filtration and lyophilization, converted 16-SAc, OTf to its water-soluble Cl salt in 90% yield.<sup>10</sup> Treatment of the latter with deoxygenated 3<u>N</u> aq. HCl (N<sub>2</sub> atm, 80°, 1 hr), lyophilization and recrystallization ( $CH_2Cl_2/ether$ ) gave 80% of 16-SH, Cl, mp 82-84°, > 95% free SH (Ellman's reagent<sup>12</sup>),<sup>13</sup> cf., eq. (1).

$$R - N(CH_3)_2 CH_2 CH_2 SCCH_3, X^{-} \xrightarrow{3N HC1} 16-SH, C1$$
  
VII (16-SAc) IV

Cleavage of PNPA by micellar 16-SH<sup>14</sup> was followed by stopped-flow spectrometry at 400 nm and pH 7.0, while nmr experiments demonstrated the sole formation (> 80%) of 16-SAc and p-nitrophenoxide. Variation of [16-SH] over 7 concentrations between  $9.24 \times 10^{-4}$  and  $3.02 \times 10^{-2} M$  gave a (pseudo-first-order) rate constant-[surfactant] profile, Fig. 1, from which  $\underline{k}_{\psi}^{\text{max}}=2.16\pm0.02_3$  sec<sup>-1</sup> at [16-SH]=0.0150M (point C, Fig. 1). At pH 7.96, k was 9.71 sec<sup>-1</sup> with 0.020M 16-SH. In Table I, the latter value is compared with analogous rate constants for the cleavage of PNPA by other micellar reagents. For comparison,  $\underline{k}_{\psi}$  for PNPA cleavage by 0.020<u>M</u> Me<sub>3</sub>NCH<sub>2</sub>CH<sub>2</sub>SH, Br (prepared by HBr cleavage of commercially available acetylthiocholine bromide) was  $0.0197+0.00196_3$  sec<sup>-1</sup> at pH 8.0 with [thiocholine]=0.0198M. The micellar thiocholine reagent is thus 493 times more reactive than the model compound under comparable conditions.

From the concentration dependence of  $\underline{k}_{ib}$  at lower concentrations of 16-SH (Gilford model 250 spectrophotometer), the "kinetic" critical micelle concentration (cmc) of 16-SH at pH  $7.0^{14}$  is ~4.2x10<sup>-4</sup>M (point A, Fig. 1).<sup>15,28</sup>

The pH dependence of  $\underline{k}_{th}$  was studied in reactions of  $4x10^{-3}M$  16-SH with  $2x10^{-5}M$  PNPA,  $\mu$ = 0.05, 23°, employing various buffers at pH 5.00, 6.17, 7.00, 7.97, 9.55, and 9.84. A plot of log  $(\underline{k}_{ij})$  vs. pH gave a sharp break-point at pH 7.3, which we take as the pK<sub>a</sub> of micellar 16-SH.<sup>18</sup> For comparison, two independent determinations of pKa for Me<sub>3</sub>NCH<sub>2</sub>CH<sub>2</sub>SH afforded 7.81 (30°) to 7.95 (20<sup>0</sup>)<sup>19a</sup> and 7.80 (temp. not reported).<sup>19b</sup>

From comparisons with other functional micellar reagents (Table I), other surfactant-thiol systems, <sup>1-3,5</sup> and thiocholine, 16-SH emerges as an extraordinarily reactive micellar reagent.<sup>20</sup> It is more reactive than other previously synthesized, self-contained, functional surfactants on the PNPA scale,<sup>21</sup> and relative to the appropriate non-micellar model, displays a greater reactivity enhancement than do other surfactant-thiol systems. 1-3,5 Indeed, toward PNPA, 16-SH  $(\underline{k}_{cat}^{=144}, pH 7.0, 23^{\circ})$  is about as reactive as ficin  $(\underline{k}_{cat}^{-173}, pH 6.9, 29.6^{\circ}).^{23}$ 

Micellization<sup>14</sup> lowers the pK<sub>a</sub> of 16-SH (7.3) relative to that of non-micellar Me<sub>3</sub>NCH<sub>2</sub>CH<sub>2</sub>SH (~7.85<sup>19</sup>), such that 29% of the 493-fold greater reactivity of the micellar thiocholine surfactant is attributable to the acid-strengthening effect of micellization and the consequent increase in the concentration of reactive thiolate ions.<sup>24</sup> The remaining catalytic advantage, a factor of 353, must have other origins. The extensive ionization of micellar 16-SH (i.e., zwitterion formation) at pH 8<sup>24</sup> results in substantial "internal" charge neutralization at the micellar surface; the concomitant hydrophobicity and accompanying desolvation may account for much of the residual enhanced reactivity<sup>22</sup> of micellar 16-SH.

Micellar 16-SH is ~3600 times more reactive than its choline analog, 16-OH (Table I,  $\frac{k_{cat}}{k_{cat}}$ scale), for which estimated  $pK_a$ 's range from 10.5<sup>25</sup> to 12.4.<sup>26</sup> Obviously, much of the catalytic advantage of the thiocholine surfactant micelles derives from greater ionization; just how much of the advantage can be attributed to factors other than acidity differences cannot be determined in the absence of a definitive  $pK_a$  for 16-OH.<sup>27</sup> We are continuing our studies of 16-SH.

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Figure 1. Pseudo-first-order rate constants for the pH 7 cleavage of PNPA by 16-SH vs. [16-SH]. Point A is the kinetic cmc of 16-SH, and point C is taken as  $k_{\mu}^{max}$  for reaction of PNPA with micellar 16-SH. Points B and B' represent identical data plotted on two different scales.

Catalyst	$\frac{k^{\max}_{\psi}}{\psi}$ (sec <sup>-1</sup> ) b	<u>k</u> cat (1/mol-sec) <sup>C</sup>	k <sup>rel</sup> cat	Ref.
CTAC1 <sup>d</sup>	0.00019 [1.35]	0.014	1.0	6
16-0H <sup>e</sup>	0.00190 [1.4]	0.136	9.7	7
AS-His-Boc	0.029 [1.6]	1.8	130.	6
16-Im <sup>e</sup>	0.20 [4.0]	5.0	360.	7
AS-Cys	1.04 [4.0]	26.0	1860.	5
16-SH	9.71 <sup>f</sup> [2.0]	485.	34600.	g

TABLE I. Cleavage of PNPA Catalyzed by Surfactant Micelles<sup>a</sup>

<sup>a</sup>See text for catalyst structures; all counterions are chloride. The reaction pH was 8.0; see ref. 14 for other conditions. <sup>b</sup>Values in [] are concentrations (<u>Mx100</u>) at which  $\underline{k}_{\psi}^{max}$  was determined.  $\frac{c_k}{E_{cat}} = \underline{k}_{\psi}^{max} / [surfactant]$ . <sup>d</sup>Cetyltrimethylammonium chloride. <sup>e</sup>0.01<u>M</u> PO<sub>4</sub> buffer. <sup>f</sup>pH 7.96. <sup>g</sup>This work.

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- Professor U. Tonellato, University of Padua, has independently prepared 16-SH, and will sepa-(8) rately report the synthetic and kinetic studies carried out in his laboratory. We thank Professor Tonellato for exchanges of correspondence.
- (9) Nmr indicated that the CH<sub>2</sub>OH multiplet of 16-OH (centered at  $\delta$ 4.07, CDCl<sub>3</sub>) was quantitatively replaced by the corresponding multiplet of VI at  $\delta 5.07$ .
- (10) Structurally consistent ir and nmr spectra, and a satisfactory elemental analysis were obtained.
- (11) The self-phase transfer reaction of 16-OTf and thioacetate is an example of a more general procedure for the synthesis of functional surfactants: R. A. Moss and W. J. Sanders, submitted for publication.
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- $CH_2N(CH_3)_2CH_2CH_2SH]$ ; the 3H S-Ac singlet of VII ( $\delta 2.40$ ) was absent in the spectrum of IV.
- (14) Conditions: 0.02<u>M</u> N<sub>2</sub>-purged PO<sub>4</sub> buffer, μ=0.05, 23<sup>o</sup>, [PNPA]=2x10<sup>-5</sup>M.
- (15) For the 6 high concentration points of Fig. 1, 16-SH with 72% free SH<sup>13,16</sup> was used, except that >95% free-SH material was employed for point A. For low concentration runs, 16-SH with 55-79% Ellman activity was used in 4 cases; material with >72% activity was used in 4 other experiments. In our experience, all samples of 16-SH with activity >55% gave comparable kinetic results with PNPA, as long as 16-SH was micellar and in substantial excess.
- (16) Despite precautions, micellar 16-SH at pH  $\geq 7$  rapidly loses activity, presumably by oxidative dimerization.  $^{17}$
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- (18) Details will appear in our full paper. Experiments with pH >7 employed the stopped-flow spectrometer with double-strength solutions of 16-SH and PNPA at pH 1-2 (several drops of HCl) in syringe A, reacted with NaOH-doped double strength buffer (syringe B); the final pH was determined after reaction.
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- (20) Analysis of the data in Fig. 1 by methods analogous to those employed in enzymatic catalysis (cf., J. H. Fendler and E. J. Fendler, "Catalysis in Micellar and Macromolecular Systems," Academic Press, New York, 1975, pp. 86ff.) affords  $k_m = 2.53 \text{ sec}^{-1}$  and K/N=31.4 for 16-SH + PNPA at pH 7. As anticipated,  $\underline{k_m} \sim \underline{k_m} a_{\overline{k}}$ ; assuming an aggregation number, N, of ~60, the binding constant, K~0.52, indicating relatively poor binding of PNPA to 16-SH micelles.
- (21) This includes AS-Cys,<sup>5</sup> as well as n-dodecyl-(2-hydroxyimidophenethyl)dimethylammonium bromide.<sup>22</sup>
- (22) T. Kunitake, S. Shinkai, and Y. Okahata, Bull. Chem. Soc. Japan, 49, 540 (1976).
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- (24) From the pKa's, micellar 16-SH is  $-\overline{83}\%$ , and the model thiocholine is -59% ionized at pH 8.
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- (27) Choosing  $pK_{a}$   $\sim 10.5$  for 16-OH, ionization would be  $\sim 0.3\%$  at pH 8. This affords a maximum nonacidity-based catalytic advantage for 16-SH of (0.3/83x3600)~13.
- (28) The facile oxidative dimerization of 16-SH precludes simple cmc determinations by conventional static procedures (i.e., surface tension).

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